Transport of Optically Active Particles from the Surface Mixed Layer: Losses Due to Grazing and Focculation During the Chalk-Ex Study

Hans G. Dam
Department of Marine Sciences
University of Connecticut
1080 Shennecossett Rd.
Groton, CT 06340-6048

Phone: (860) 405-9098 Fax: (860) 405-9153 (Fax) e-mail: hans.dam@uconn.edu

George B. McManus Department of Marine Sciences University of Connecticut Groton, CT 06340-6048

Phone: (860) 405-9164 Fax: (860) 405-9153 (Fax) e-mail: george.mcmanus@uconn.edu

Award Number: N000140110016 http://www.marinesciences.uconn.edu/faculty/dam.html

LONG-TERMS GOALS

To determine the mass balance of optically active particles within the surface boundary layer and to identify processes responsible for their redistribution.

OBJECTIVES

- 1) To perform manipulative experiments in which a known quantity of optically active CaCO₃ particles are introduced into the surface mixed layer, and tracked over time and space. This approach effectively removes uncertainty in the production term of the mass balance equation.
- 2) To quantify the loss from the mixed layer of optically active particles due to grazing and aggregation.

APPROACH

In addition to the work by Dam and McManus, there is close collaboration with Drs. W.M. Balch and C. Pilskaln (Bigelow Lab for Ocean Sciences/Optical and vertical flux studies) and Dr. A. Pluddemann (WHOI/ physical studies). Their work is not included in this report.

In November 2001, we participated in a 10-day cruise in the Gulf of Maine and the slope waters in the Western North Atlantic Ocean. During the cruise, we participated in the deployment of two chalk patches and conducted bottle experiments to investigate the fate of the chalk with regard to processes such as grazing and aggregation. In addition to this field component, laboratory observations on chalk/grazer interactions are ongoing.

maintaining the data needed, and of including suggestions for reducing	lection of information is estimated to completing and reviewing the collect this burden, to Washington Headqu uld be aware that notwithstanding ar DMB control number.	ion of information. Send comments a arters Services, Directorate for Infor	regarding this burden estimate of mation Operations and Reports	or any other aspect of the 1215 Jefferson Davis	nis collection of information, Highway, Suite 1204, Arlington
1. REPORT DATE 30 SEP 2002 2. REPORT TYPE			3. DATES COVERED 00-00-2002 to 00-00-2002		
4. TITLE AND SUBTITLE				5a. CONTRACT NUMBER	
Transport of Optically Active Particles from the Surface Mixed Layer: Losses Due to Grazing and Focculation During the Chalk-Ex Study				5b. GRANT NUMBER	
Losses Due to Grazing and Focculation During the Chark-Ex Study				5c. PROGRAM ELEMENT NUMBER	
6. AUTHOR(S)				5d. PROJECT NUMBER	
				5e. TASK NUMBER	
				5f. WORK UNIT NUMBER	
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Department of Marine Sciences, University of Connecticut, 1080 Shennecossett Rd., Groton, CT, 06340				8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES)				10. SPONSOR/MONITOR'S ACRONYM(S)	
				11. SPONSOR/MONITOR'S REPORT NUMBER(S)	
12. DISTRIBUTION/AVAILABILITY STATEMENT Approved for public release; distribution unlimited					
13. SUPPLEMENTARY NOTES					
	nass balance of option		within the surfa	ce boundary	layer and to
15. SUBJECT TERMS					
16. SECURITY CLASSIFIC	17. LIMITATION OF ABSTRACT	18. NUMBER OF PAGES	19a. NAME OF RESPONSIBLE PERSON		
a. REPORT unclassified	b. ABSTRACT unclassified	c. THIS PAGE unclassified	Same as Report (SAR)	6	ACCIONALE I ENSON

Report Documentation Page

Form Approved OMB No. 0704-0188

WORK COMPLETED

Grazing. During the Chalk-Ex cruise in November, 2001, we conducted five bottle incubations on deck while the artificial chalk patches were being followed. Experiments consisted of four treatments. For all treatments, one-liter polycarbonate bottles were filled with water pre-screened through 200 μm mesh by reverse flow. Three of the treatments received added mesozooplankton in successively greater multiples of the natural abundance, up to c. 32x. The experiments were designed to look for evidence of microzooplankton grazing (removal of chalk-sized particles in the <200 μm treatment) and either top-down control by mesozooplankton (grazing on chalk-sized particles declines as mesozooplankton concentration increases) or direct mesozooplankton grazing (grazing on chalk-sized particles increases with mesozooplankton concentration). Abundance of chalk-sized particles (1.3-3.5 μm) was measured with an Elzone particle counter at initial and final (c. 20h) experimental times. We also had formalin-killed controls, but they showed evidence of particle production during the experiments, most likely due to mineral precipitation of the preservative's buffer, and were thus difficult to interpret.

Flocculation. During the same cruise we measured the flocculation rates of particles assemblages, from waters in the chalk patches, in Couette devices (Drapeau et al., 1994, Dam and Drapeau, 1995) by monitoring the changes in mean particle size with time. Measurements were done on five separate occasions in triplicate incubations lasting 5-6 h each. The shear rate in the Coutte devices was 6 s⁻¹. We employed two kinds of devices to measure particle size—an electric impedance device (EIZONE 280 particle counter) and a laser counter (GALAI). Both of these devices can resolve particles as small as 0.5 μm. The chalk particle size ranged from 1.3 to 3.5 μm.

RESULTS

Grazing. Net per capita rates of change for chalk-sized particles ranged from –0.89 to +0.47 d⁻¹ in the <200 μm treatment during the five experiments (fig. 1). Three of the five experiments showed evidence of top-down control on microzooplankton grazing. Because the chalk size-fraction also contained coccoid cyanobacteria and heterotrophic bacterioplankton, which presumably were growing during the incubations, any net losses would provide minimal estimates of potential grazing on chalk. For several experiments, chalk particles were abundant enough to count directly using a polarized light microscope. These observations suggested real net loss of chalk particles in the bottles. Because processes other than grazing might have contributed to loss of chalk particles in the bottles (e.g. dissolution or aggregation). We plan future work on possible chalk ingestion by protists using laboratory cultures. Preliminary observations with a large heterotrich ciliate, *Condylostoma sp.*, in laboratory culture, indicate that this species will ingest chalk.

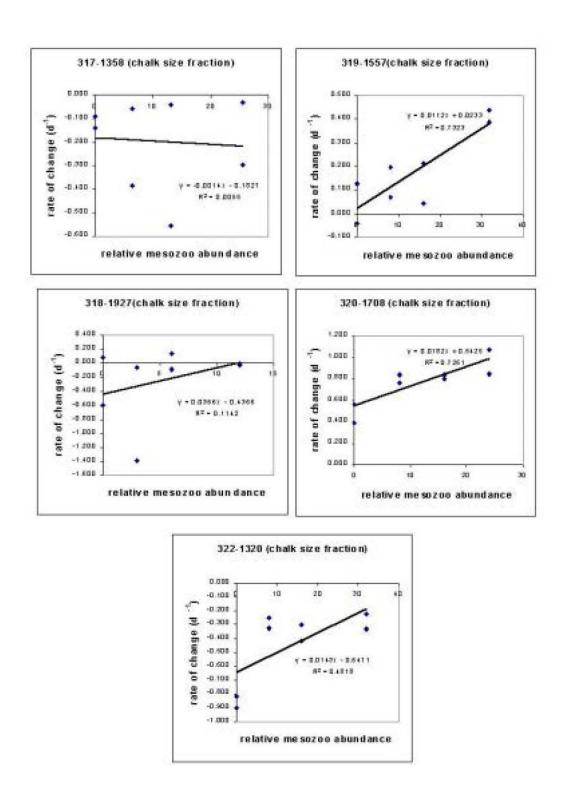


Figure 1. Plots of mesozooplankton abundance vs. net rate of change of chalk-sized particles (1-3 µm) in the five bottle experiments. The value at zero on the x-axis indicates the rate of change due to microzooplankton (<200 µm). As the relative abundance of mesozooplankton increases, the rate of change should decrease if these larger grazers are ingesting the chalk, or increase if they are instead primarily grazing on the microzooplankton that are ingesting the chalk.

Flocculation. Our experiments on particle flocculation were predicated on the assumption that volume is conserved during aggregation. Hence, in a closed system (as in the Couette devices) as aggregation takes place, mean particle size increases with time. We observed very good agreement between both instruments in characterizing particle size during the incubations. Furthermore, we also observed particle size increase with time during all of our five separate incubations, as illustrated in the particle size spectra shown in Fig. 2. That is, the slope of the line for each spectrum decreased with time. These changes in spectra translate to a 2.5–fold increase in mean particle volume during the length of the 5-6 h incubation. However, it is also evident that volume was not conserved during the incubations since the Y-intersect of the spectra increased with time. This was characteristic of all five incubations during the cruise. Two possible explanations account for the lack of volume conservation in the incubations. One is that aggregates were fractal. The other is that new particles were formed during the incubations, either from bacterial or phytoplankton growth, or from aggregation of colloids. In any case, the results obtained here pose a challenge to characterizing the chalk aggregation with classical flocculation theory.

318-1927 (Elzone) frequency distribution

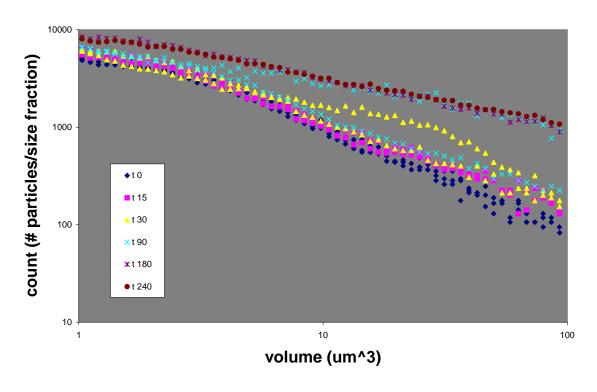


Figure 2. Mean particle volume (X-axis) versus particle abundance (counts/0.2 ml) for each size fraction (Y-axis) as a function of time. The slope of each line decreases with time indicating aggregate formation during the incubations. The Y-intersect for each line also increases with time, indicating that volume is not conserved during the incubations.

IMPACT/APPLICATIONS

Our experiments are designed to identify the relative importance of two loss terms—grazing and aggregation—on the mass balance of coccoliths, an important group of optically active particles. Without this information the evolution of the underwater field and prediction of underwater visibility on the spatial (1-10,000m horizontal and 1-100 m vertical) and temporal scales (hours to days) of coccolithophore blooms is severely hindered.

TRANSITIONS

None.

RELATED PROJECTS

We work as a part of a team with the other PI's in this project: Drs. Balch and Pilskaln (Bigelow Laboratory for Ocean Sciences) and Dr. Pluddemann (WHOI).

REFERENCES

Dam, H.G. and D.T. Drapeau. 1995. Coagulation efficiency, organic-matter glues and dynamics of particles during a phytoplankton bloom in a mesocosm study. Deep-Sea Res. II. 42: 111-123.

Drapeau, D.T., H.G. Dam and G. Grenier. 1994. An improved flocculator design for use in particle aggregation experiments. Limnol. Oceanogr. 39: 723-729.